

## Glipizide and Metformin Hydrochloride Tablets

» Glipizide and Metformin Hydrochloride Tablets contain not less than 90.0 percent and not more than 110.0 percent of the labeled amounts of glipizide ( $C_{21}H_{27}N_5O_4S$ ) and metformin hydrochloride ( $C_4H_{11}N_5 \cdot HCl$ ).

**Packaging and storage**—Preserve in well-closed containers, and store at controlled room temperature.

### Add the following:

• **Labeling**—When more than one *Dissolution* test is given, the labeling states the *Dissolution* test used only if *Test 1* is not used. • (RB 1-Dec-2009)

**USP Reference standards** <11>—*USP Glipizide RS*. *USP Glipizide Related Compound A RS*. *USP Metformin Hydrochloride RS*. *USP Metformin Related Compound A RS*.

### Identification—

#### A: GLIPIZIDE—

**Infrared Absorption** (197A)—Prepare the test specimen as follows. Transfer not fewer than 10 Tablets to a suitable container, add 10 mL of methanol, and shake to remove any tablet coating. Drain the methanol, add 20 mL of water, and stir until the Tablets dissolve (about 1 hour). Transfer the solution to a separatory funnel, and extract twice with 10-mL portions of chloroform, shaking for approximately 5 minutes. Transfer the lower organic layer into a beaker containing 3 to 4 g of anhydrous magnesium sulfate. Repeat the extraction of the solution in the separatory funnel two more times, each time using 20-mL portions of chloroform. Swirl the mixture in the beaker for about 1 minute. Filter, and collect the filtrate. Evaporate the solvent under vacuum, and dry the residue under vacuum for 4 hours at 105°. Mound the residue onto a diamond cell: the IR spectrum so obtained exhibits maxima only at the same wavelengths as a similarly obtained spectrum of USP Glipizide RS.

#### B: GLIPIZIDE—

The retention time of the major peak in the chromatogram of the *Assay preparation* corresponds to that in the chromatogram of the *Standard preparation*, as obtained in the *Assay for glipizide*.

#### C: METFORMIN HYDROCHLORIDE—

The retention time of the major peak in the chromatogram of the *Assay preparation* corresponds to that in the chromatogram of the *Standard preparation*, as obtained in the *Assay for metformin hydrochloride*.

### Change to read:

### Dissolution <711>—

• TEST 1—• (RB 1-Dec-2009)

**Medium:** 0.05 M phosphate buffer, pH  $6.8 \pm 0.05$  (Prepared by dissolving 12.96 g of monobasic potassium phosphate and 1.66 g of sodium hydroxide in approximately 400 mL of water, and diluting with water to 2000 mL. Adjust the pH, if necessary, with diluted sodium hydroxide. [NOTE—Tight control of the pH is critical.]); 1000 mL.

**Apparatus 2:** 50 rpm.

**Times:** 45 minutes for glipizide, 30 minutes for metformin hydrochloride.

Determine the amount of glipizide ( $C_{21}H_{27}N_5O_4S$ ) dissolved by employing the following method.

**Buffer solution**—Dissolve approximately 3.4 g of monobasic potassium phosphate in approximately 800 mL of water. Adjust with

10 N sodium hydroxide solution to a pH of  $6.0 \pm 0.1$ . Dilute with water to 1000 mL, and mix.

**Mobile phase**—Prepare a filtered and degassed mixture of methanol and *Buffer solution* (13 : 12). Make adjustments if necessary (see *System Suitability* under *Chromatography* <621>).

**Standard solution**—Transfer about 50 mg of USP Glipizide RS, accurately weighed, to a 1000-mL low-actinic volumetric flask, and dissolve in 100 mL of methanol. Dilute with *Medium* to volume, and sonicate for about 5 minutes. [NOTE—This solution is stable for 7 days at 5° when protected from light.]

**Working standard solution**—Dilute the *Standard solution* with *Medium* in order to obtain a solution containing L/1000 mg per mL, with L being the glipizide tablet label claim, in mg.

**Test solution**—After the specified time, withdraw about 10 mL of the solution under test. Pass the solution through a 0.45- $\mu$ m PVDF filter or a 1.0- $\mu$ m glass fiber filter, discarding the first mL.

**Chromatographic system** (see *Chromatography* <621>)—The liquid chromatograph is equipped with a 220-nm detector and a 4.6-mm  $\times$  15-cm column that contains 5- $\mu$ m packing L1. The flow rate is about 1.0 mL per minute. Chromatograph the *Working standard solution*, and record the peak responses as directed for *Procedure*: the relative standard deviation for replicate injections is not more than 2.0%.

**Procedure**—Separately inject equal volumes (about 50  $\mu$ L) of the *Working standard solution* and the *Test solution* into the chromatograph, record the chromatograms running for about 8 minutes, and measure the peak response for glipizide. Calculate the amount of glipizide ( $C_{21}H_{27}N_5O_4S$ ) dissolved by the formula:

$$\frac{r_U \times C_S \times 1000 \times 100}{r_S \times L}$$

in which  $r_U$  and  $r_S$  are the peak responses obtained from the *Test solution* and the *Working standard solution*, respectively;  $C_S$  is the concentration, in mg per mL, of glipizide in the *Working standard solution*; 1000 is the volume, in mL, of *Medium*; 100 is the conversion factor to percentage; and L is the tablet label claim for glipizide, in mg.

Determine the amount of metformin hydrochloride ( $C_4H_{11}N_5 \cdot HCl$ ) dissolved by employing the following method.

**Test solution**—After the specified time, withdraw about 10 mL of the solution under test. Pass the solution through a 0.45- $\mu$ m PVDF filter or a 1.0- $\mu$ m glass fiber filter, discarding the first mL.

**Procedure**—Determine the amount of metformin hydrochloride dissolved by employing UV absorption at the wavelength of maximum absorbance at about 233 nm on portions of the *Test solution*, suitably diluted with *Medium*, if necessary, in comparison with a *Standard solution* having a known concentration of USP Metformin Hydrochloride RS in the same *Medium*. Calculate the amount of metformin hydrochloride ( $C_4H_{11}N_5 \cdot HCl$ ) dissolved by the formula:

$$\frac{A_U \times C_S \times 1000 \times 100}{A_S \times L}$$

in which  $A_U$  and  $A_S$  are the absorbances obtained from the *Test solution* and the *Working standard solution*, respectively;  $C_S$  is the concentration, in mg per mL, of the *Working standard solution*; 1000 is the volume, in mL, of *Medium*; 100 is the conversion factor to percentage; and L is the tablet label claim for metformin hydrochloride, in mg.

**Tolerances**—Not less than 80% (Q) of the labeled amount of  $C_{21}H_{27}N_5O_4S$  is dissolved in 45 minutes. Not less than 80% (Q) of the labeled amount of  $C_4H_{11}N_5 \cdot HCl$  is dissolved in 30 minutes.

• TEST 2—If the product complies with this test, the labeling indicates that the product meets USP *Dissolution Test 2*.

## 2 Glipizide

**Medium:** 0.05 M phosphate buffer, pH 6.8; 1000 mL.

**Apparatus 2:** 50 rpm.

**Time:** 60 minutes for both metformin hydrochloride and glipizide.

**pH 6.0 Buffer solution**—Dissolve 4.3 g of octanesulfonic acid sodium salt and 6.9 g of monobasic monohydrate sodium phosphate in 1000 mL of water. Adjust with diluted sodium hydroxide to a pH of  $6.00 \pm 0.05$ .

**Glipizide standard stock solution**—Transfer about 25 mg of USP Glipizide RS, accurately weighed, to a 500 mL volumetric flask. Dissolve in and dilute with methanol to volume.

**Working standard solution**—Transfer an accurately weighed quantity of USP Metformin Hydrochloride RS to a volumetric flask, add a suitable aliquot of *Glipizide standard stock solution*, and dilute with *Medium* to obtain a final concentration of L/1000 mg per mL, where L is the tablet label claim for both metformin hydrochloride and glipizide, in mg.

**Test solution**—Pass a portion of the solution under test through a suitable 0.45- $\mu$ m polyethersulfone filter, discarding the first few mL.

**Mobile phase**—Prepare a filtered and degassed mixture of methanol and pH 6.0 Buffer solution (1 : 1). Make adjustments if necessary (see *System Suitability* under *Chromatography* (621)).

**Chromatographic system** (see *Chromatography* (621))—The liquid chromatograph is equipped with a 260-nm detector, a sample compartment chiller maintained at 4°, and a 4.6-mm  $\times$  25-cm column that contains packing L1. The flow rate is about 1.0 mL per minute. Chromatograph the *Working standard solution*, and record the peak responses as directed for *Procedure*: the resolution, *R*, between glipizide and metformin hydrochloride is not less than 2, and the relative standard deviation for replicate injections is not more than 2.0% for both glipizide and metformin hydrochloride.

**Procedure**—Separately inject equal volumes (about 20  $\mu$ L) of the *Working standard solution* and the *Test solution* into the chromatograph, record the chromatograms, and measure the peak response for both glipizide and metformin hydrochloride. Calculate the percentage of glipizide and metformin hydrochloride dissolved by the formula:

$$\frac{r_U \times C_S \times 1000 \times 100}{r_S \times L}$$

in which  $r_U$  and  $r_S$  are the peak responses obtained for glipizide or metformin hydrochloride from the *Test solution* and the *Working standard solution*, respectively;  $C_S$  is the concentration, in mg per mL, of glipizide or metformin hydrochloride in the *Working standard solution*; 1000 is the volume, in mL, of *Medium*; 100 is the conversion factor to percentage; and *L* is the tablet label claim for glipizide or metformin hydrochloride, in mg.

**Tolerances**—Not less than 80% (*Q*) of the labeled amount of  $C_{21}H_{27}N_5O_4S$  and of  $C_4H_{11}N_5 \cdot HCl$  is dissolved in 60 minutes. (RB 1- Dec-2009)

**Uniformity of dosage units** (905): meet the requirements.

### Related compounds—

GLIPIZIDE—

**Ammonium phosphate buffer, Solution A, Solution B, and Chromatographic system**—Prepare as directed in the *Assay for glipizide*.

**Test solution**—Use the *Assay preparation*, prepared as directed in the *Assay for glipizide*.

**Procedure**—Inject about 50  $\mu$ L of the *Test solution* into the chromatograph, record the chromatogram, and measure the peak responses. Calculate the percentage of glipizide related compound A

(approximate relative retention time 0.92) and other individual impurities in the portion of Tablets taken by the formula:

$$100(1/F)(r_i / r_s)$$

in which *F* is the relative response factor for each impurity and is equal to 1.4 for glipizide related compound A and 1.0 for all other peaks;  $r_i$  is the peak response of each impurity; and  $r_s$  is the sum of the responses of all the peaks: not more than 2.0% of glipizide related compound A is found; not more than 0.5% of any other individual glipizide related impurity (eluting after approximately 8 minutes) is found; and not more than 1.0% total impurities, excluding glipizide related compound A, is found. [NOTE—Disregard the broad peak due to metformin that elutes before 8 minutes. Disregard any peak observed in the blank, and disregard any peak less than 0.05%.]

### Chromatographic purity—

METFORMIN HYDROCHLORIDE—

**Solution A, Solution B, Mobile phase, and Chromatographic system**—Prepare as directed in the *Assay for metformin hydrochloride*.

**Test solution**—Use the *Assay preparation*, prepared as directed in the *Assay for metformin hydrochloride*.

**Procedure**—Inject about 25  $\mu$ L of the *Test solution* into the chromatograph, record the chromatogram, and measure the peak responses. Calculate the percentage of each impurity in the portion of Tablets taken by the formula:

$$100(r_i / r_s)$$

in which  $r_i$  is the peak response for each impurity, and  $r_s$  is the sum of the responses of all the peaks: not more than 0.1% of any individual impurity is found; and not more than 0.5% of total impurities is found. [NOTE—Disregard any peak less than 0.05%, and disregard any peak observed in the blank.]

### Assay for glipizide—

**Ammonium phosphate buffer**—Dissolve 2.6 g of dibasic ammonium phosphate in water, and dilute with water to 1000 mL. Adjust with ammonium hydroxide to a pH of 8.0.

**Solution A**—Prepare a degassed mixture of water, *Ammonium phosphate buffer*, and acetonitrile (14 : 5 : 1).

**Solution B**—Prepare a degassed mixture of acetonitrile, *Ammonium phosphate buffer*, and water (2 : 1 : 1).

**Diluent**—Use a mixture of acetonitrile and water (60 : 40).

**Standard stock preparation**—Transfer an accurately weighed quantity of USP Glipizide RS to a suitable low-actinic volumetric flask. Dissolve first in acetonitrile, using 60% of the final volume, by sonicating for about 20 minutes, then dilute with water to volume to obtain a solution having a known concentration of about 0.1 mg of glipizide per mL. [NOTE—The solution is stable for 2 weeks when stored at 5° protected from light.]

**Standard preparation**—Transfer 25.0 mL of *Standard stock preparation* to a 200-mL low-actinic volumetric flask. Dilute first with 75 mL of *Diluent*, and bring to volume with water to obtain a solution having a known glipizide concentration of approximately 0.0125 mg per mL. [NOTE—The solution is stable for 2 weeks when stored at 5° protected from light.]

**System suitability preparation**—Transfer approximately 5 mg of USP Glipizide Related Compound A RS to a 500-mL volumetric flask, and fill halfway with acetonitrile. Sonicate for about 30 minutes to dissolve, and dilute with acetonitrile to volume. Transfer 1 mL of this solution to a 50-mL low-actinic volumetric flask, and dilute with *Standard preparation* to volume.

**Assay preparation**—Transfer not fewer than 5 Tablets to a suitable volumetric flask, and fill halfway with *Diluent*. Sonicate for 30 minutes, and shake vigorously for another 30 minutes to dissolve. Dilute with water to volume, and mix to obtain a solution with a final glipizide concentration of about 0.0125 mg per mL. Pass a

portion of this solution through a nylon or PVDF filter having a 0.2- $\mu$ m porosity, and use the filtrate. [NOTE—The solution is stable for 2 weeks when stored at 5° protected from light.]

**Chromatographic system** (see *Chromatography* <621>)—The liquid chromatograph is equipped with a 223-nm detector and a 4.6-mm  $\times$  15-cm column that contains 5- $\mu$ m packing L7. The flow rate is about 1.0 mL per minute. The chromatograph is programmed as follows.

Time (minutes)	Solution A (%)	Solution B (%)	Elution
0–3	100	0	isocratic
3–18	100→0	0→100	linear gradient
18–20	0	100	isocratic
20–22	0→100	100→0	linear gradient
22–30	100	0	re-equilibration

Chromatograph the *System suitability preparation* and the *Standard preparation*, and record the peak responses as directed for *Procedure*. For the *System suitability preparation*, the relative retention times are about 0.92 for glipizide related compound A and 1.0 for glipizide; the resolution, *R*, between glipizide related compound A and glipizide is greater than 1.2. For the *Standard preparation*, the relative standard deviation for replicate injections is less than 2.0%.

**Procedure**—Separately inject equal volumes (about 50  $\mu$ L) of the *Standard preparation* and the *Assay preparation* into the chromatograph, record the chromatograms, and measure the responses for the glipizide peaks. Calculate the quantity, in mg per Tablet, of glipizide ( $C_{21}H_{27}N_5O_4S$ ) by the formula:

$$CV(r_U / r_S)/N$$

in which *C* is the concentration, in mg per mL, of USP Glipizide RS in the *Standard preparation*; *V* is the volume, in mL, of the *Assay preparation*; *r<sub>U</sub>* and *r<sub>S</sub>* are the glipizide peak responses obtained from the *Assay preparation* and the *Standard preparation*, respectively; and *N* is the number of Tablets taken to prepare the *Assay preparation*.

#### Assay for metformin hydrochloride—

**Solution A**—Prepare a 50 mM hexanesulfonic acid solution by dissolving 9.41 g of sodium 1-hexanesulfonate in 1000 mL of water, and adjusting with trifluoroacetic acid to a pH of 2.0.

**Solution B**—Prepare a solution of water and acetonitrile (60 : 40).

**Mobile phase**—Prepare a degassed mixture of water, *Solution A*, and *Solution B* (50 : 30 : 20). Make adjustments if necessary (see *System Suitability* under *Chromatography* <621>).

**Diluent pH 2.0**—Prepare a mixture of water, *Solution A*, and acetonitrile (63 : 30 : 7).

**Standard preparation**—Dissolve an accurately weighed quantity of USP Metformin Hydrochloride RS in *Diluent pH 2.0* to obtain a solution having a known concentration of about 0.1 mg per mL.

**System suitability preparation**—Dissolve a suitable quantity of USP Metformin Related Compound A RS in water to obtain a solution containing about 5  $\mu$ g per mL. Pipet 0.5 mL of this solution into a 50-mL volumetric flask, and dilute with the *Standard preparation* to volume.

**Assay preparation**—Quantitatively dilute a portion of the *Assay preparation*, obtained as directed in the *Assay for glipizide*, with *Diluent pH 2.0*, to obtain a solution having an expected concentration of about 0.1 mg of metformin hydrochloride per mL, based on the label claim.

**Chromatographic system** (see *Chromatography* <621>)—The liquid chromatograph is equipped with a 218-nm detector and a 4.6-mm  $\times$  15-cm column that contains 3.5- $\mu$ m packing L11. The flow rate is about 1.0 mL per minute. Chromatograph the *System suitability preparation*, and record the peak responses as directed for *Procedure*: the relative retention times are about 0.26 for metformin related compound A and 1.0 for metformin; the resolution, *R*, between the two peaks is not less than 3.0; and the relative standard deviation for replicate injections, determined from the metformin peak, is less than 2.0%.

**Procedure**—Separately inject equal volumes (about 25  $\mu$ L) of the *Standard preparation* and the *Assay preparation* into the chromatograph, record the chromatograms, and measure the responses for the major peaks. Calculate the quantity, in mg per Tablet, of metformin hydrochloride ( $C_4H_{11}N_5 \cdot HCl$ ) by the formula:

$$CVD(r_U / r_S)/N$$

in which *C* is the concentration, in mg per mL, of USP Metformin Hydrochloride RS in the *Standard preparation*; *V* is the volume, in mL, of the *Assay preparation*, as prepared in the *Assay for glipizide*; *D* is the dilution factor of the *Assay preparation*; *r<sub>U</sub>* and *r<sub>S</sub>* are the peak responses obtained from the *Assay preparation* and the *Standard preparation*, respectively; and *N* is the number of Tablets used to prepare the *Assay preparation*.